Cell Death And Zika Virus: An Integrated Network Of The Mechanisms Of Cell Injury

Jorge Rodrigues de Sousa1,2
Raimunda do Socorro da Silva Azevedo1
Juarez Antônio Simões Quesma2–4
Pedro Fernando da Costa Vasconcelos1,4

1Departamento de Arbovirologia e Febres Hemorrágicas, Instituto Evandro Chagas, Ananindeua, Pará, Brazil;
2Núcleo de Medicina Tropical Belém, Universidade Federal do Pará, Belém, Pará, Brazil;
3Departamento de Patologia, Instituto Evandro Chagas, Ananindeua, Pará, Brazil; 4Universidade do Estado do Pará, Belém, Pará, Brazil

Correspondence: Pedro Fernando da Costa Vasconcelos
Evandro Chagas Institute, Rodovia BR-316, km-07, s/n, Ananindeua 67030-000, Brazil
Email pedrovasconcelos@iec.gov.br

Abstract: Zika virus (ZIKV) is an arbovirus that is transmitted by Aedes mosquitoes. Its prototype was isolated in 1947 from serum of a sentinel Rhesus monkey (Macaca mulatta) in the Zika forest of Uganda. As a member of the genus Flavivirus, family Flaviviridae, ZIKV is enveloped and icosahedral and possesses a single-stranded, positive-sense RNA genome of approximately 10.7 kb. Epidemiologically, infection by ZIKV has become a global health concern in recent years because of the occurrence of epidemics, its speed of dissemination, routes of transmission, and the sequelae it can cause especially in newborns. At the neural level, there are still many gaps in our understanding of the mechanisms that induce ZIKV infection-associated microcephaly. However, some studies already demonstrated that underlying cell death is determinant to induce the congenital malformation. In this report, we reviewed the various mechanisms of cell injury involved in the immunopathogenesis of ZIKV infection and discussed its relationship with the death of neuronal and glial cells development and microcephaly.

Keywords: ZIKV, microcephaly, neuroinflammatory response, cell death

Perspectives
Zika virus (ZIKV) is a member of the family Flaviviridae and the genus Flavivirus; it was originally isolated in 1947 from a febrile sentinel rhesus monkey in the Zika forest (Uganda).1 In 2015, the World Health Organization (WHO) declared the Zika epidemic a public health emergency in Brazil owing to the outbreak and exponential growth of reported cases of microcephaly and other congenital defects.1,2

ZIKV exhibits extreme versatility and modulates mechanisms of immune evasion; recent experimental advances have shown that ZIKV infects different cell types in several organs and tissues, confirming that transmission occurs not only by bites of infected mosquitoes (Aedes), but also by sexual and vertical routes.3 In this context, it is believed that receptors belonging to the TIM and TAM families, particularly AXL, facilitate ZIKV entry in cells.4

Considering the concept of immune privilege and the high selectivity of the blood-placental, blood-testis, and blood–brain barrier to control protein and immune cell entry in the organs, the immune responses mediated by dendritic cells, M1/M2 macrophages, endothelial cells, and CD4 and CD8 lymphocytes reflect the complexity of the host defense system and its organization to combat infectious agents.5–7

Recent experimental studies in vitro and in vivo have shown that several mechanisms of cell injury are involved in the immunopathogenesis of ZIKV infection. These studies suggest that there is a link between microcephaly and...
cell death resulting from the neuroinflammatory response, whereby apoptosis, necrosis, necroptosis, pyroptosis, paraptosis, and autophagy increase the death of neuronal and glial cells and, consequently, result in brain atrophy and ZIKV-induced microcephaly.

**Apoptosis, Necrosis, And Necroptosis**

The development of microcephaly presents a great challenge owing to the lack of a comprehensive understanding of the evolutionary mechanisms and dynamics of the process. Accordingly, further studies are needed to clarify these issues and to determine the mechanisms underlying cell injury. A loss of tissue homeostasis and thereby a cell functional imbalance can result in the induction of necrosis or apoptosis pathways.

Recent more investigations showed the involvement of components of the necroptotic cell death pathway to limit neuronal Zika virus infection. In cortical neurons, the activation of Z-DNA-binding protein 1 (ZBP1) and RIPK1/RIPK3 can trigger the formation of intracellular cascade which modulate the expression of immunoresponsive gene 1 (IRG1), as well as itaconate metabolite. In this process, itaconate inhibits the formation of succinate dehydrogenase (SDH) and consequently suppresses the replication of viral genomes.

With respect to apoptosis, extrinsic and intrinsic pathways act jointly to enhance cell damage. In the extrinsic pathway, cytokines, growth factors, and death ligands induce the activation of death receptors responsible for initiate the intrinsic cascade. Activation of the intrinsic cascade results in mitochondrial damage that alters the electron transportation chain, provokes the formation of pores in the membrane where cytochrome C is released to bind with Apaf1, and forms the apoptosome complex responsible for activating a series of caspases that direct the cell to apoptosis. This process involves the caspases 6, 7, 8, 9, and, in particular, caspase 3, which mediate programmed cell death induced by ZIKV. Classic apoptosis leads to a process of natural cell death in which the host defense system induces various mechanisms to maintain homeostasis, without causing major damage; in microcephaly cases, apoptosis induces the uncontrolled death of neurons and glia cells at several stages.

This has direct relationship with cellular cycle as evidenced in study cell death showing that in the ventricular zone, progression and mitotic abnormalities of progenitor neural cells in the brain are associated with the intense production of caspase 3, 7, 8, and 9.

**Pyroptosis**

Dynamic, interacting processes are linked to the immune response, and recent study of cell death mechanisms has shown that endoplasmic reticulum stress directly contributes to the activation of signaling proteins that modulate excitotoxicity, increase intracellular calcium production, and enhance cell damage. In the CNS, an intense neuroinflammatory response and loss of mitochondrial homeostasis results in the development of a new cell death mechanism that is extremely harmful to neurons and glial cells, i.e., pyroptosis in microcephaly cases ZIKV-induced.

Interestingly, pyroptosis differs from apoptosis and necrosis since it is dependent on the activation of the inflammasome, a multiprotein complex characterized by the activation of NLRP3 and AIM2, receptors that recognize pathogen-associated molecular patterns (PAMPs) of the ZIKV and induce the release of caspase 1 to activate pyroptosis and trigger the maturation of IL-1β, IL-18, and IL-33. Studies of these cytokines are necessary to understand the neuroinflammatory mechanisms involved in the development of microcephaly; IL-1β, IL-18, and IL-33 belong to the IL-1 family and modulate the activities of the M1 (IL-1β and IL-18) and M2 (IL-33) macrophages/microglia as well as the Th1 (IL-1β and IL-18) and Th2 (IL-33) lymphocytes.

Recently in experimental in vitro was showed that ZIKV enhance proinflammatory response to promote NLRP3 inflammasome activation by stabilizing caspase-1 to suppress cGAS-mediated type I IFN signaling. In this context, NS1 block the proteasomal degradation of the caspase 1 recruiting USP8 to cleave K11-linked poly-ubiquitin chains from caspase-1 at Lys134. The implications of this process enhance inflammasome activation and inhibit type I IFN signaling.

**Paraptosis**

As an immune escape mechanism and adaptation, ZIKV uses the cellular machinery to transform the cell into a setting for viral replication, inducing massive cytoplasmic vacuolization in human epithelial cells, primary human fibroblasts, and human astrocytes. This process is associated with a new form of cell death that does not result in apoptotic corpsules, known as paraptosis. The mechanisms underlying paraptosis in cells of the CNS involve PIK3, AKt, PERK, IRE-1, and ATF-6, as well as vesicle transport orchestrated by the endoplasmic reticulum and Golgi.
Interesting that ZIKV-induced vacuoles are dependent on viral translocation into ER through Sec61 and on PI3K/Akt signaling. Therefore, the paraptosis makes the cell a dangerous ZIKV reservoir via the massive cytoplasmic vacuolization, functional alterations of the endoplasmic reticulum, delayed phagolysosome maturation, and the excessive production of ROS; accordingly, it can contribute to cell destruction and ZIKV dissemination.

**Autophagy**

In addition to paraptosis, another cell death pathway implicated in the pathogenesis of ZIKV-infection is autophagy. Autophagy regulation in the brain is associated with ZIKV tropism by several CNS cells. In vitro the infection of human fetal neural stem cells (fNSCs) with ZIKV leads to impaired neurosphere formation and elevated autophagy. Infection with ZIKV strains MR766 and IbH30656 efficiently induced LC3-I to LC3-II conversion and LC3 puncta formation of fNSCs in the presence or absence of lysosome inhibitor bafilomycin A1. A highlight is that in this process two ZIKV proteins (NS4A and NS4B) inhibit AKt and consequently the PIK3 cascade and mTOR production causing the deregulation of neurogenesis aberrant activation of autophagy. Curiously both proteins produced by dengue virus do not have the same effect on neurogenesis. Therefore, the identification of this mechanism suggests a strong actuation of NS4A and NS4B to modulate the immune evasion of ZIKV in the viral pathogenesis.

**Conclusion And Future Direction**

Many questions about microcephaly induced by ZIKV remain. However, investigations of several mechanisms of cell injury involved in the immunopathogenesis of ZIKV infection suggest that there is a link between microcephaly and cell death in which the neuroinflammatory response in situ can induce massive cytoplasmic vacuolization, delayed phagolysosome maturation, and the excessive production of ROS. Consequently, it can contribute to cell destruction and ZIKV dissemination.
apoptosis, necrosis, and pyroptosis and consequently result in brain atrophy. In vitro triggering of the necroptosis, paraptosis, and autophagy enhance death of neuronal and glial cells. Therefore, identification of these mechanisms enables the discovery of several immune-targets for anti-ZIKV therapeutic intervention. However, we emphasize that studies in vivo need to be performed to confirm this hypothesis. In summary, we propose an integrated model that synthesizes the relationships among these underlying mechanisms (Figure 1).

**Funding**

This work was supported by the Ministry of Science, Technology and Innovation/National Council for Scientific and Technological Development CNPQ/Brazil (grant numbers: 303999/2016-0, 439971/2016-0, and 440405/2016-5), and CAPES (Zika Fast-track).

**Author contributions**

All authors contributed towards data analysis, drafting and critically revising the paper, gave final approval of the version to be published, and agreed to be accountable for all aspects of the work.

**Disclosure**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**References**
